

## Biology and Management of Septoria Spot of Citrus

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Septoria spot of citrus caused by *Septoria citri* occurs mainly on mature, cold-injured fruit in rainy winter seasons. Due to recent outbreaks, Septoria spot has become a quarantine disease in some export countries for California citrus fruit. Fruit marketed to these destinations have to be certified as disease-free.

Characteristic signs of *S. citri* (i.e., fruiting bodies – pycnidia, conidia) on fruit lesions are often not present, and thus the disease is difficult to diagnose. In addition, the fungus also grows very slowly, and *S. citri* colonies are often overgrown by other fungi occurring on citrus. Therefore, we developed a new molecular detection assay.

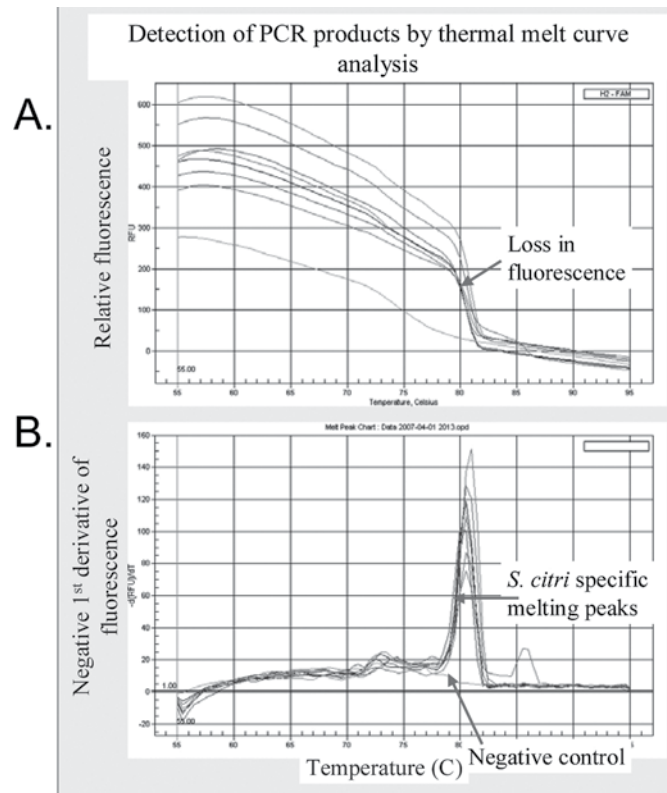


Figure 1. Thermal melt curve analysis of PCR products using *Septoria citri*-specific DNA primers. A. Melt curve relative fluorescence over a range of temperatures for 8 samples. B. Negative first derivative of fluorescence melt curve shown in A.

PCR primers derived from the beta-tubulin gene were designed that specifically amplify *S. citri* DNA in extracts of citrus fruit. Because of the risk of amplicon contamination in a diagnostic laboratory, PCR products are detected non-electrophoretically by thermal melt curve analysis immediately after amplification. PCR and melt curve analysis are done in a single set-up in a real-time thermocycler.

In thermal melt curve analysis of PCR products, the thermal stability of DNA duplexes is measured and graphed as a melt curve (Fig. 1A). The fluorescent dye SYBR Green that is incorporated into the PCR reaction binds to double-stranded DNA molecules. A loss of fluorescence is observed when the dye dissociates from the separating DNA strands at a specific temperature. The negative first derivative of fluorescence is then automatically graphed against a temperature scale and a melt curve is displayed (Fig. 1B). Melt temperatures are specific for different PCR products.

Thus, in samples positive for *S. citri*, specific DNA fragments are produced in PCR amplifications, and their presence is indicated by a peak at a specific melt temperature in a diagram. For verification of DNA quality and to exclude false negatives, a primer pair specific for orange DNA (provided by Dr. M. Roose, UC Riverside) is also used in PCR amplifications. This primer pair produces a second peak at a different melt temperature. The absence of this specific peak in a sample using these orange-specific primers indicates the sample that lacks DNA or has non-amplifiable DNA.

Using both pathogen- and orange- specific PCR primers - with each PCR product having a unique temperature melt curve - a single-step automated procedure was developed to assess suspect lesions for Septoria spot (Fig. 2).

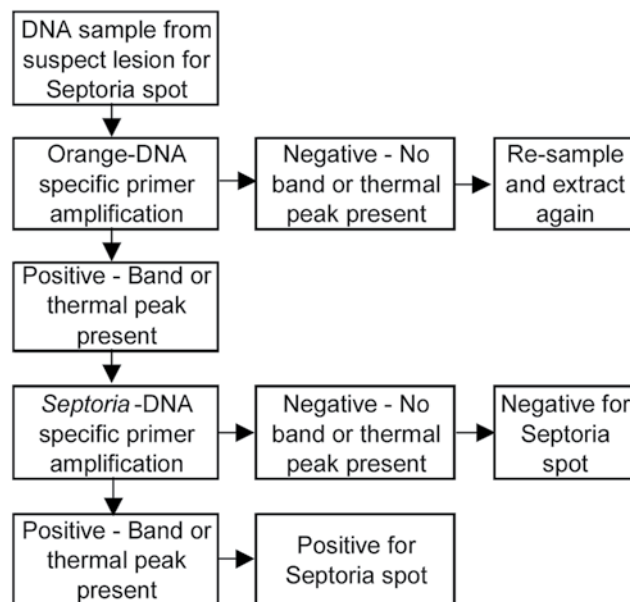


Figure 2. Flow chart for automation of PCR detection of *Septoria spot* of citrus.

This new PCR method was extensively evaluated using orange fruit from the NAVEK (Navel and Valencia Exports to Korea) incubation program at the Kearney Agricultural Center. Samples positive for *Septoria spot* in fungal isolations were always positive in the PCR assay, whereas samples infected with a range of other fungi never showed a specific melt peak in the assay. Thus, thermal melt curve analysis reliably and rapidly detects *S. citri* in infected citrus fruit, and the procedure will be setting a standard for the detection of this quarantine pathogen.

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